Background: Using garlic is widespread in Iran and other countries as a medicine and a natural spice. Garlic is a potential inhibitor for food pathogens. Foods contaminated with pathogens pose a potential danger to the consumer's health. The use of garlic can increase the shelf life and decrease the possibilities of food poisoning and spoilage in processed foods.

Objectives: The aim of this study was to investigate the antibacterial effect of garlic aqueous extract on growth of Staphylococcus aureus bacteria.

Materials and Methods: In this study, the garlic aqueous extract was prepared under sterile conditions and was added in 1, 2, and 3 mL to 100g hamburger samples. A group of samples was prepared to be used as treatment sample, while a group was stored at 4°C and -18°C. The samples were kept in refrigerator for one and two weeks and they were frozen for one, two and three months and then subjected to microbial tests.

Results: Statistical evaluation of the first and second week samples indicated a significant growth decreased by all the 1, 2, and 3-mL extracts. In treatment of one, two and three-month samples, the growth of S. aureus was significantly decreased by the 2 and 3-mL extracts. The 1-mL extract was effective in decreasing the growth, and a significant difference was observed in treatments with 2 and 3-mL extracts. However, there was no significant difference between the two and three-month samples, though they were significantly different from the one-month samples. After evaluations, treatment with the 2-mL extract was found to be the best one.

Conclusions: Garlic aqueous extract has antibacterial properties against S. aureus present in hamburger. Moreover, garlic aqueous extract can be used not only as a flavor but also as a natural additive for hamburger. In addition, garlic has antibacterial properties against other Gram-positive and Gram-negative bacteria, which must be investigated in further studies.

Keywords: Antibacterial Effect; Garlic; Staphylococcus aureus; Hamburger

1. Background

Consumption of red meat and meat products has been dramatically increased all over the world. Among foods, hamburger is a very popular ready-to-eat and/or ready-to-cook meal as it is delicious and enjoyable with an easy consumption method plus being prepared with or without added salt and seasonings (1, 2). A retail sale of fast food hamburgers has turned into a big business. Americans consume nearly five billion hamburgers each year, equal to 1.2 billion pounds of meat (3). Lipid oxidation and growth of microbes during the maintenance period are regarded as two important factors, affecting the quality of this food material, and are among the concerns of hamburger producers, since they may have negative effects on the odor and taste of hamburger, consequently decreasing the number of consumers (1, 4). Although application of antioxidinants and antibacterials with chemical origins are useful for maintaining the quality, increased maintenance duration, prevention from lipid oxidation, and economic loss can have adverse effect on health of this product and by virtue of standards, using such materials in hamburger is prohibited.

One of the reasons for the consumers' high interest in hamburger rather than other meat products such as sausages is lack of chemical additives in hamburger, which increases the health level of this product. Pathogens related to food poisoning, which can be isolated from foods with animal origins such as meat and meat products (like hamburger), are specific types of bacteria called Salmonella serotypes, Staphylococcus aureus, Listeria monocytogenes and Escherichia coli O157: H7 (1, 2, 5, 6). They can be transferred to the water surface of carcasses when cattle is slaughtered (6). This highlights the necessity of studying the application of natural additives such as garlic. Garlic is a main component of hamburger which is used...
to improve its odor and taste. Moreover, it has numerous medicinal usages and treatment properties, but rarely considered as a healthy food additive (7).

Garlic (Allium sativum L.) belongs to the Liliaceae family and falls within the group of onion, like mountain onion and shallot, and also has antibacterial activity against many common pathogenic bacteria (8, 9). Cavallito and Bailey (10) studied the chemical properties of garlic for the first time. They tried a host of methods for producing garlic extract and finally managed to produce the most important garlic compounds, called allicin, diallyl disulfide and alipin, which have less odor. They believed that extracted allicin, if used with ethyl alcohol solvent, would have more stability and antibacterial effects (10, 11). Among the organosulfur ingredients of garlic, S-allyl cysteine sulf oxide or alliin is the odorless compound. When garlic is cut, it is influenced by the alliinase enzyme, which is the cysteine sulfoxide lyase, and turns into allicin. All these substances are responsible for the strong smell of garlic and have antibacterial and antioxidant properties (12).

Allicin is the main biologically and biochemically active component of freshly crushed garlic which can easily be decomposed under the influence of heat and time into stable compounds. 2-propenesulfenic acid and tioacrolein are two important molecules derived from garlic, which in turn result in the formation of greater molecules including diallyl trisulfide and dithiin (13, 14). Generally, garlic is a potential antibiotic and shows an effective performance against the bacteria resistant to pharmaceutical antibiotics. In 1970’s, Europeans tested the garlic extract on 10 species resistant to bacteria and yeasts and found it very resistant to Salmonella, S. aureus, Mycobacterium and Proteus species (15). In the present study, The antibacterial effects of garlic aqueous extract on 133 species of bacteria (both Gram-positive and Gram-negative) resistant to medicine, including S. epidermidis, S. aureus, Streptococcus pneumoniae, S. pyogenes, Haemophilus influenzae, S. typhi, Pseudomonas aeruginosa, E. coli, Shigella spp., Proteus spp. and 10 types of candida were studied. The minimum inhibition concentrations (MIC) or lowest densities of garlic aqueous extract that prevented the growth of microorganisms were determined in the present study (Table 1) (16, 17).

2. Objectives

This study aimed to investigate the antibacterial effects of garlic aqueous extract on S. aureus growth prevention in hamburger under the freezing condition, during one and two weeks as well as one, two, and three months.

3. Materials and Methods

3.1. Preparing the Garlic Aqueous Extract

Fresh garlic was purchased from a local market in Hamadan city, Iran, and 500 g of that was washed, peeled and washed again with sterile distilled water. Then, it was blended and crushed with 200 mL cool physiology serum (0.9%) on top of chopped ice, next to fire flame to supply the sterilized condition. The obtained mixture was sterilized with an ordinary colander and then it was thinned with a flat, sterile 0.2-μl syringe and stored at -20°C (18, 19).

3.2. Hamburger Preparation

Samples of hamburger were produced in accordance with the Institute of Standards and Industrial Research of Iran, Number 2304, under laboratory conditions. The raw materials were prepared from food stores and formulation of samples was performed under sterile conditions. The purchased grinded meat was blended on top of ice under sterile conditions with the colandered onion. Salt and common spicy foods along with flour in a specified ratio were blended with meat. Extracts of 1, 2, and 3 mL were added to the samples. The prepared hamburgers were put into refrigerator in 100-g moulds and then the test for detection and counting S. aureus was performed with three repeats (20).

3.3. Detection and Counting Staphylococcus aureus

During the detection stage, a suspension of the hamburger treatments was prepared and about 1-2 mL of the first dilution (10^1) was transferred to the test tube containing cooked meat culture medium with 10% additional salt (S. aureus are halophilic bacteria). Afterwards, it was incubated at 37°C for 48 hours. A high percentage of salt contributed to the growth of S. aureus and if they were damaged during the process they could be repaired. Next, the incubated mixture was transferred to Baird-Parker Agar (BPA) where linear incubation was performed in a loop. S. aureus at incubation created black colonies with transparent areola after heating. To count S. aureus, after preparing the first dilution (10^1) and homogenizing 10 g of hamburger and 90 mL of sterile diluents (physiological serum), we tried to make further dilutions up to 10^-8 to 10^-10. The dilutions, cultured through surface method, were distinguished and the colonies containing S. aureus in 1 g of the sample were counted and evaluated (12, 21).

3.4. Sensational Evaluation

The formulated hamburgers were studied in laboratory by hedonic method with respect to color, appearance, odor, taste, crispiness, and structure. Sensational evaluation was performed by 15 panelists who studied

<table>
<thead>
<tr>
<th>Type of Microorganism</th>
<th>MIC, mg/mL</th>
</tr>
</thead>
<tbody>
<tr>
<td>Gram-positive bacteria</td>
<td>15.6-48.3</td>
</tr>
<tr>
<td>Gram-Negative bacteria</td>
<td>22.9-37.2</td>
</tr>
<tr>
<td>Molds and yeasts</td>
<td>14.9-15.5</td>
</tr>
</tbody>
</table>

Abbreviation: MIC, minimum inhibitory concentration.
all the mentioned factors in the indicator sample as well as in the samples with garlic aqueous extracts of 1, 2, 3 mL, and finally categorized them as excellent, very good, good, acceptable and unacceptable levels. In this test, sample A was used as an indicator sample, while samples B, C and D contained 1, 2 and 3 mL garlic aqueous extracts, respectively. Then, the results were statistically studied based on complete random block plans (22, 23).

3.5. Statistical Analysis

Statistical evaluation was performed using SPSS software and the average treatments were compared using the Duncan’s test.

4. Results

Results of studying the effects of garlic aqueous extract on growth of *S. aureus* in hamburger sample are presented in Tables 2 and 3 Figure 1. The *S. aureus* count in the hamburger sample after the test and on zero days (immediately after adding garlic extract) was $2.6 \times 10^6$ CFU/g. In the control group, *S. aureus* growth in the second week of the treatment significantly increased compared with the same group’s value in the first week. Meanwhile, the 1, 2, and 3-mL treatments had no significant difference for the first week and two weeks, and all three concentrations of garlic liquid extract prevented the bacteria from growing and multiplying over time.

Growth of *S. aureus* in one and two-week samples indicated that all the extracts of 1, 2, and 3 mL significantly decreased the growth of *S. aureus*, with the effect of 3 mL extract being higher than the 1 and 2-mL extracts. However, considering a probable error of 5%, there was no significant difference in effects of the extracts. The 2 and 3-mL extracts showed a significant difference when applied to the indicator sample, while application of the 1-mL extract did not. Statistical studying of the one, two and three-month samples indicated that 2 and 3-mL extracts with the probable error of 5% significantly decrease the *S. aureus* count, with no significant difference. The 1-mL extract was effective on decreasing the bacterial growth and showed a significant difference with the 2 and 3-mL extracts. There was no significant difference between the results from the two and three-month samples and they were significantly different from the results of the one-month sample, so that less *S. aureus* growth was observed in the one-month sample.

The storage methods comparison also revealed the fact that freezing is an excellent way to preserve meat products because meat has a flexible fiber structure which prevents from serious damages to tissues against freezing and reduces the growth of microorganisms. However, freezing does not completely remove the microorganisms and it can only reduce their process and functionality.

### Table 2. *Staphylococcus aureus* in One and Two-Week Hamburger Samples

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Indicator Treatment</th>
<th>1-mL Treatment</th>
<th>2-mL Treatment</th>
<th>3-mL Treatment</th>
<th>Standard</th>
</tr>
</thead>
<tbody>
<tr>
<td>Counted bacteria at $10^5$, first week</td>
<td>$2 \pm 1^{b,c}$</td>
<td>$6.66 \pm 2.3^{c}$</td>
<td>$0.00 \pm 0.00^{b}$</td>
<td>$1 \pm 1.73^{b}$</td>
<td>$10^3$</td>
</tr>
<tr>
<td>Counted bacteria at $10^5$, second week</td>
<td>$14.33 \pm 13.05^{c}$</td>
<td>$6.33 \pm 6.5^{c,b}$</td>
<td>$0.66 \pm 1.15^{b}$</td>
<td>$0.00 \pm 0.00^{b}$</td>
<td>$10^3$</td>
</tr>
</tbody>
</table>

*Data are presented as mean ± SD. The number of bacteria (CFU/g) is indicated in 1 g of contaminated food. To calculate the numbers it should be multiplied by $10^6$. Sample A. Sample B.*

### Table 3. *Staphylococcus aureus* in One, Two, and Three-Month Hamburger Samples

<table>
<thead>
<tr>
<th>Treatments</th>
<th>Indicator Treatment</th>
<th>1-mL Treatment</th>
<th>2-mL Treatment</th>
<th>3-mL Treatment</th>
<th>Standard</th>
</tr>
</thead>
<tbody>
<tr>
<td>Counted bacteria at $10^3$, first month</td>
<td>$1 \pm 1.73^{b}$</td>
<td>$0.33 \pm 0.57^{b}$</td>
<td>$1 \pm 1^{b}$</td>
<td>$0.33 \pm 0.57^{b}$</td>
<td>$10^3$</td>
</tr>
<tr>
<td>Counted bacteria at $10^3$, 2nd month</td>
<td>$121.33 \pm 23.79^{c}$</td>
<td>$95.67 \pm 9.71^{d}$</td>
<td>$5.67 \pm 4.04^{b}$</td>
<td>$4.33 \pm 1.52^{b}$</td>
<td>$10^3$</td>
</tr>
<tr>
<td>Counted bacteria at $10^3$, 3rd month</td>
<td>$120.33 \pm 13.5^{c}$</td>
<td>$111.67 \pm 11.59^{d}$</td>
<td>$3 \pm 2.64^{b}$</td>
<td>$2.67 \pm 1.52^{b}$</td>
<td>$10^3$</td>
</tr>
</tbody>
</table>

*Data are presented as mean ± SD. The Number of Bacteria is indicated (CFU/g) in 1 g of Contaminated Food. To calculate the numbers they should be multiplied by $10^4$. Sample A. Sample B. Sample C.*
A. sativum L. has a membrane lipid content. Aqueous extracts showed growth in all these three treatments. The maximum growth was observed by the third extract concentration which is three times more effective on Gram-positive bacteria than Gram-negative ones. Such results will be accounted for the membrane lipid content of these two bacteria which helps the easy penetration of allicin into the membrane and consequently influences the RNA (12).

It can be concluded that garlic aqueous extract has antibacterial properties against S. aureus present in hamburger. Moreover, garlic aqueous extract can be used not only as a flavor but also as a natural additive for hamburger. Garlic also has antibacterial properties against other Gram-positive and Gram-negative bacteria, which should be investigated in further studies.

5. Discussion

S. aureus is a Gram-positive, catalase-positive, aerobic and non-aerobic, halophilic, and immobile bacterium, capable of manitol fermentation. This bacterium has an irregular cluster shape and is a member of the Micrococcus family and yeast under glucose aerobic conditions, and is killed at over 62°C (pasteurization temperature), with the ability to tolerate this temperature at least half an hour. Existence of golden S. aureus in food is regarded to as a potential danger which results in food poisons, due to existence of a nauseant polypeptide called enterotoxin (21, 24). Researches have indicated S. aureus as a very sensitive bacterium to aqueous and alcoholic extracts. Fujisawa et al. (14) reported chemical and biological stability of garlic aqueous and alcoholic extracts and their effects on both Gram-positive (S. aureus) and Gram-negative (E. coli) bacteria.

It was found that alcoholic garlic extract was much more stable and more effective than the aqueous extract. However, tests revealed that the antimicrobial activity of garlic is totally dependent on the allicin compound, which is three times more effective on Gram-positive bacteria than Gram-negative ones. Such results will be accounted for the membrane lipid content of these two bacteria because E. coli has a membrane lipid content 10 times higher than S. aureus, making allicin unable to reach the goal, as they are trapped in this lipid content. In previous surveys by Deresse (12) on the antibacterial effects of garlic extracts on preventing the growth of S. aureus in vitro, 30 different species of S. aureus were isolated and the Ethiopian garlic (A. sativum L.) aqueous extracts effects were studied on them.

It was indicated that MIC and garlic aqueous extract shall be more than 7.5 mg/mL to prevent bacterial growth. It also became clear that if we had autoclaved the garlic aqueous extract under the temperature of 121°C for 15 minutes, it would lose its efficiency on any strains of S. aureus and would not be effective anymore. Notably, antimicrobial properties of raw garlic under room temperature and garlic kept at -10°C were similar and the fresh garlic had grater effects. Studying the antimicrobial properties of garlic indicated that garlic is full of anion compounds including nitrates, chlorides, sulfides and organosulphur compounds that can be easily resolved in water and are responsible for antibacterial properties (25). In addition, several studies on garlic have indicated that allicin in garlic is the most important substance that creates antibacterial properties and limits the speed of RNA synthesis; however, the preliminary role of allicin is trapping RNA. Lipid is a part of S. aureus membrane, which helps the easy penetration of allicin into the membrane and consequently influences the RNA (12).

The statistical evaluation of S. aureus showed that hamburger samples of the three extracts of 1, 2, and 3 mL of garlic extract had a significant effect in reducing the S. aureus growth in all these three treatments. The maximum effect was observed by the third extract concentration and the extracts with 1 and 2 mL concentrations were also effective in reducing the S. aureus growth. Furthermore, in the present study, it was indicated that 1, 2, and 3-mL garlic aqueous extracts were effective on destruction of S. aureus in hamburger, as the principal goal of garlic aqueous extract was to destroy this bacteria and prevent from performance of the protein inducing RNA synthesis.

References


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